



CONCORDANCE BETWEEN NASOPHARYNGEAL CULTURES AND PATHOGENS IN EAR DISCHARGE AND LOWER AIRWAY SPECIMENS FROM INDIGENOUS CHILDREN IN THE NORTHERN TERRITORY

Hare Kim M¹, Cheng Allen C^{1,2,3}, Smith-Vaughan Heidi¹, Chang Anne C^{1,4,5}, Morris Peter S^{1,6}, Leach Amanda J¹

1 Child Health Division, Menzies School of Health Research, Charles Darwin University, Darwin, NT;

2 Department of Epidemiology and Preventive Medicine, Monash University, Melbourne, Victoria;

3 Infectious Diseases Unit, Alfred Hospital, Melbourne, Victoria;

4 Queensland Children's Medical Research Institute, University of Queensland, Brisbane, Queensland;

5 Department of Respiratory Medicine, Royal Children's Hospital, Brisbane, Queensland;

6 Flinders Medical School, Royal Darwin Hospital, Darwin, NT.

Introduction

Why look at concordance?

- The nasopharynx (NP) is the [main] source of respiratory bacteria infecting the middle ear and lower airways
- Additionally, NP swabs are relatively easy to collect compared to ear discharge or lower airway specimens
- Prevalent invasive pneumococcal disease serotypes are not necessarily common carriage types
- This is less true for mucosal disease
- Surveillance of NP carriage provides important information as new pneumococcal conjugate vaccines (PCVs) are introduced

Methods (studies)

We examined concordance between NP bacteria and bacteria found in ear discharge (ED) and broncho-alveolar lavage (BAL) fluid from Indigenous children

Data were reviewed from studies of:

- (1) acute otitis media with perforation (AOMwiP), and
- (2) chronic suppurative otitis media (CSOM) between 1996 and 2005;
- (3) bronchiectasis in children undergoing bronchoscopy from 2007 to 2011; and
- (4) OM (not here specified) from 2008 to 2011 (study ongoing)

Methods (laboratory)

Standard published laboratory methods were used for culture and identification, except that:

- Different studies have different protocols for the number of colonies isolated per positive specimen
- This is unlikely to significantly alter the proportion of specimens found positive for each pathogen
- However it may affect the number of strains detected (e.g. pneumococcal serotypes) per positive specimen
- While ED rarely contains >1 serotype, multiple types are frequently isolated from BAL specimens

Results

Data were obtained from:

- (1) 266 ear assessments in 140 children with AOMwiP;
- (2) 179 ear assessments in 133 children with CSOM;
- (3) 104 Indigenous children with radiographically confirmed bronchiectasis*; and
- (4) 130 children with ED from at least one ear.

* Lower airway infection was defined as $>10^4$ cfu/mL BAL fluid (for any individual pathogen) to exclude possible upper airway contamination introduced during the bronchoscopy procedure

Isolation of respiratory pathogens in ear discharge and bronchoalveolar lavage from NT Indigenous children

	AOMwiP (n=266)	CSOM (n=179)	Bronchiectasis (n=104)
<i>Streptococcus pneumoniae</i>	32%	22%	16% >10 ⁴ cfu/mL BAL
<i>Haemophilus influenzae</i>	42%	46%	31% >10 ⁴ cfu/mL BAL
<i>Moraxella catarrhalis</i>	13%	25%	12% >10 ⁴ cfu/mL BAL

Sensitivity and specificity

Nasopharyngeal swabs are sensitive but not specific for pathogens found in the middle ear (previous studies)

What does this mean?

If a particular bug is found in ED from a particular child, it is likely (~85-95% certainty) that the same bug will be found in the NP culture from that child

However, finding a particular bug in NP culture is no guarantee that the same bug will be found in ED from the same child

Nevertheless, at a population level NP swabs are useful indicators of strains circulating (e.g. pneumococcal serotypes) and their antibiotic resistance

Sensitivity of NP culture for pathogens in ED and BAL fluid from NT Indigenous children

	<u>AOMwIP</u> (n=266)	CSOM (n=179)	Bronchiectasis (n=104)
<i>Streptococcus pneumoniae</i> (same serotype)	32% 94% (77%)	22% 90% (74%)	16% 71% (100%)*
<i>Haemophilus influenzae</i>	42% 90%	46% 83%	31% 81%
<i>Moraxella catarrhalis</i>	13% 85%	25% 93%	12% 92%

* BAL fluid had some additional types not found in NP swabs

Specificity of NP culture for pathogens in ED and BAL fluid from NT Indigenous children

	AOMwiP (n=266)	CSOM (n=179)	Bronchiectasis (n=104)
<i>Streptococcus pneumoniae</i>	32% 34%	22% 30%	16% 70%
<i>Haemophilus influenzae</i>	42% 26%	46% 25%	31% 64%
<i>Moraxella catarrhalis</i>	13% 14%	25% 22%	12% 76%

Higher specificities for BAL fluid due to azithromycin?
(associated with reduced NP carriage)

Sensitivity and specificity of NP culture for pathogens in BAL fluid from children who had or had not received azithromycin

	Azithromycin within 2 weeks (n=39)	No recent antibiotics (n=39)	
<i>Streptococcus pneumoniae</i>	Sens=56% (5/9) Spec=80%	Sens=100% (6/6) Spec=42%	↑ ↓
<i>Haemophilus influenzae</i>	Sens=60% (6/10) Spec=66%	Sens=92% (12/13) Spec=58%	↑ ↓
<i>Moraxella catarrhalis</i>	Sens=100% (1/1) Spec=82%	Sens=80% (4/5) Spec=62%	Outlier? ↓

Without antibiotics (compared to azithromycin), sensitivity of NP for BAL pathogens is increased and specificity decreased (i.e. more similar to ED)

MARSii data (Monitoring Antibiotic Resistance and Serotypes) from NT & WA Indigenous children

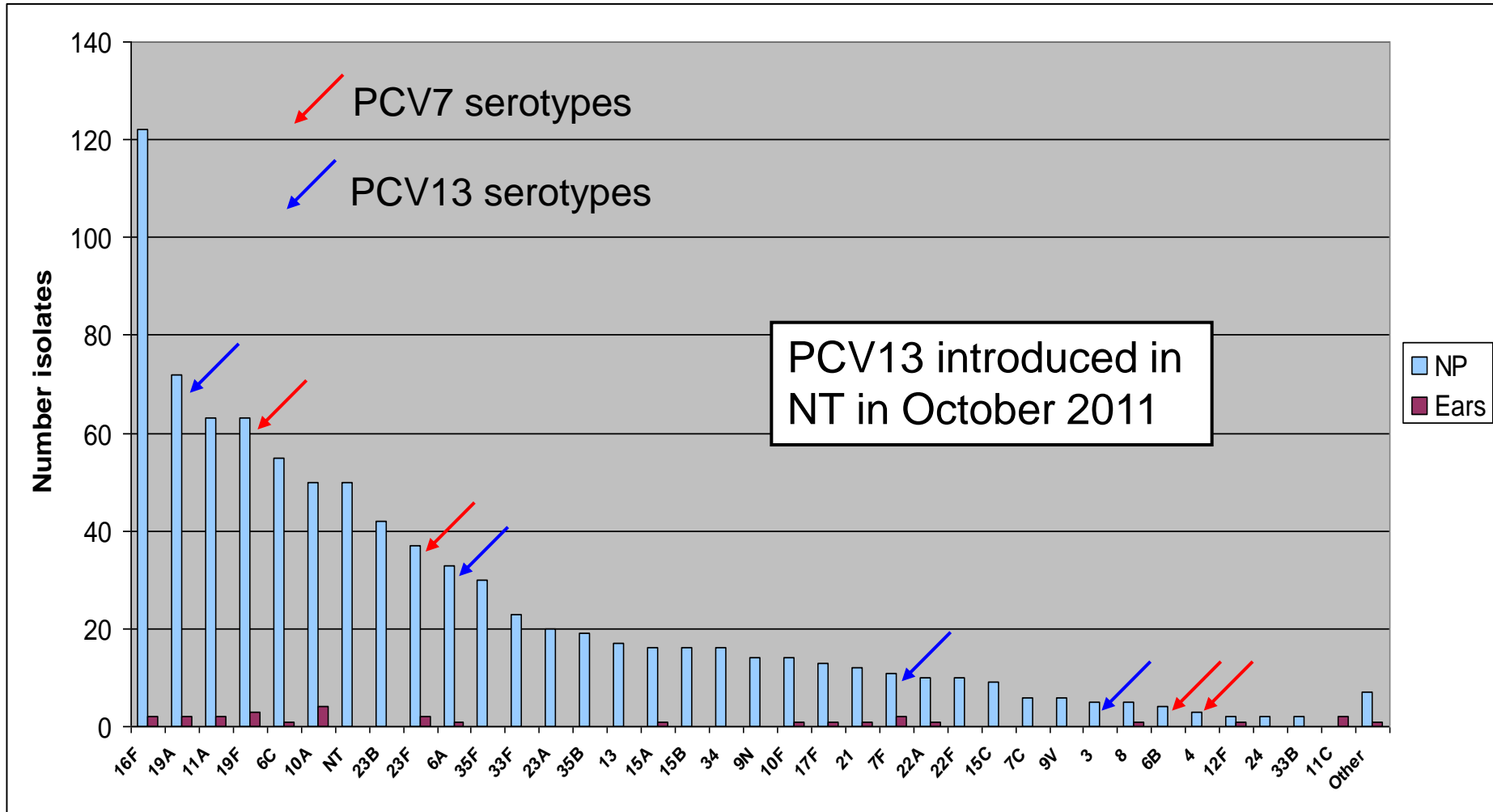
Incomplete data (study ongoing)

133 OM episodes – 3 had no matching NP swab (**n=130**)

	NP carriage	Ears	Sensitivity	Specificity
<i>Streptococcus pneumoniae</i>	81%	21%*	93%	22%
<i>Haemophilus influenzae</i>	77%	47%	80%	26%
<i>Moraxella catarrhalis</i>	48%	1.5%	na	na

* Same serotype in 90% of NP/ear pairs

Serotype hierarchy 2008-2011 (MARSi)



Conclusions

- Nasopharyngeal (NP) swabs are sensitive but not specific for pathogens found in the middle ear
- NP swabs are also sensitive, and more specific (possibly), for lower airway pathogens
- Epidemiological surveillance for pneumococcal serotypes based on NP swabs is likely to reflect serotypes found in severe OM and lower airway infection
- Further work on multiplicity may improve concordance (e.g. NP swabs preceding bronchoscopy or ear exams to identify additional serotypes or NTHi strains)
- Interventions to reduce carriage may reduce these infections

Acknowledgements

We thank:

- The children and families who participated in the studies
- Menzies clinical staff who collected the specimens
- Menzies laboratory staff who processed the specimens
- Allen Cheng who did the statistical analysis for the AOMwiP and CSOM data to 2005
- The OMOZ committee for organising this meeting

