

Otitis Media

Are probiotics an answer ?

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Outline

- Background
- Study Design
- *in vitro* Assays
- Future Directions

Otitis Media (OM)



OM – one of the most common infection in children (Rovers et al 2004)



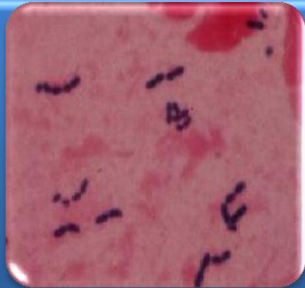
Aboriginal children – one of the highest prevalence of OM (Gunasekara et al 2008)



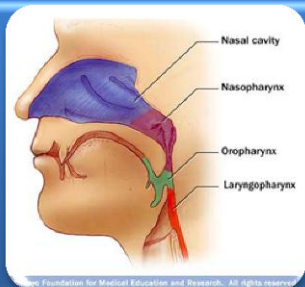
Starts as URI > acute OM > chronic OM (Kong et al 2009)



Recurrent ear discharge > hearing loss > poor speech development
(Williams et al 2009)



Streptococcus pneumoniae is one of the main bacteria causing OM (Stoodley et al 2008)



Colonise the nasopharynx > travel to middle ear > OM (Wiertsema et al 2009)

How to tackle OM ?

- **Various strategies**
- ***S. pneumoniae***
 - Early colonisation (3 m) - increased risk of OM (Faden et al 1997)
 - Indigenous children earlier (3 wks) (Smith et al 2008)
 - Increased bacterial load - increased risk of OM (Smith et al 2006)
- ***Can inhibition of colonisation be an alternative method in OM prevention?***

Probiotics

- Live micro organisms- confer health benefits to the host
- Possible mechanisms of action
 - Competitive inhibition of microbes
 - Immuno-modulation
 - Bacteriocin production
- Alpha haemolytic streptococci - inhibitory effect against OM pathogens

(Iannitti et al 2010)

(Tano et al 1999)

Streptococcus salivarius

- Potential pharyngeal probiotic
(Guglielmetti et al 2010)
- Safe, commercially available
- Produces bacteriocins, BLIS
- Harbours megaplasמידs
(Tagg et al 2009)



Does *S. salivarius* inhibit the colonisation of *S. pneumoniae* ?

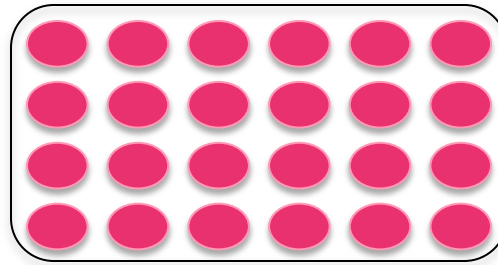
Study Design

- *in vitro* lab assays
- *in vivo* animal experiments
- Clinical trial

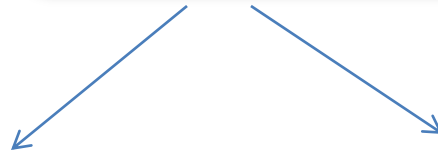
in vitro Lab Assays

- Administration of *S. salivarius* will inhibit *S. pneumoniae* colonisation of **HEp-2 cells** in an in vitro laboratory setting
- Probiotic – *S. salivarius* (Ss)
- Bacteria – *S. pneumoniae* (Pnc)
- Epithelial cell- **HEp-2cell** (Human Epidermoid laryngeal carcinoma cell)

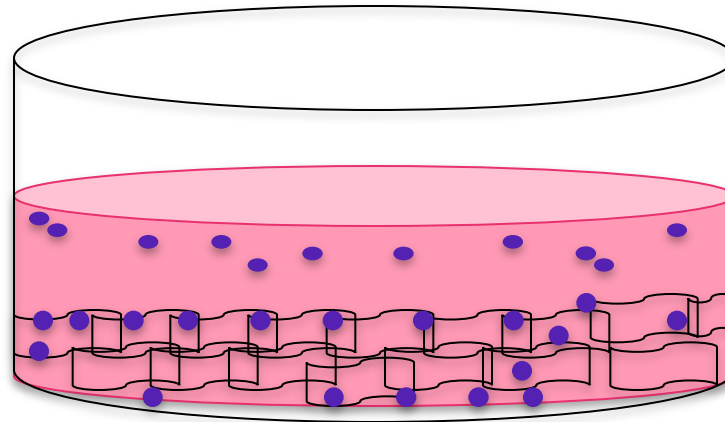
The Assay



24-well tray



Pneumococci
HEp-2 cells



Various testing conditions

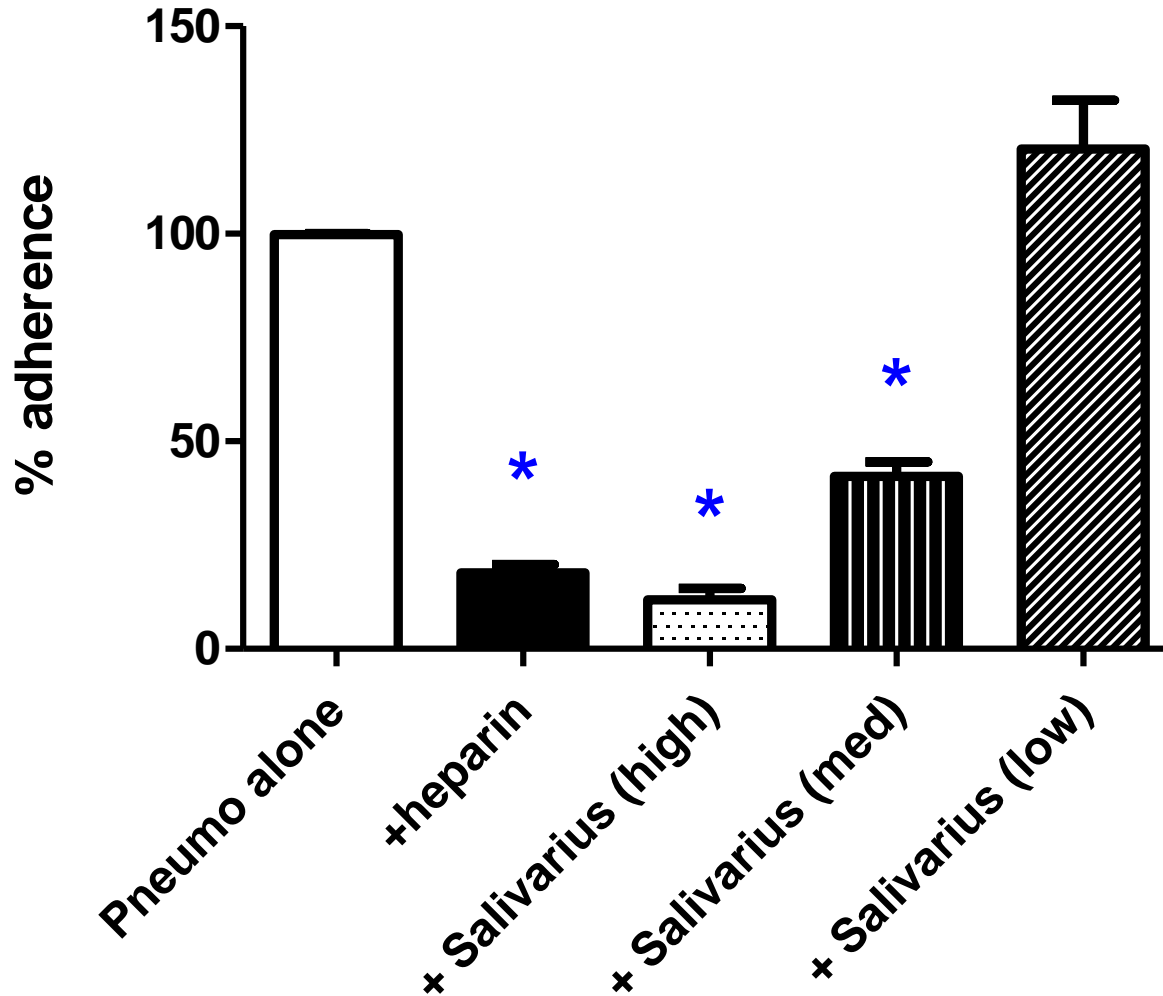
- cells & Pnc (standard)
 - cells , Pnc & heparin (control)
 - cells , Pnc & Ss (high)
 - cells , Pnc & Ss (medium)
 - cells , Pnc & Ss (low)
-
- **Quantification of Pnc - qPCR**

Addition of *S. salivarius*

- Pre-administration – one hour before Pnc
- Co-administration – along with Pnc
- Post-administration – one hour after Pnc

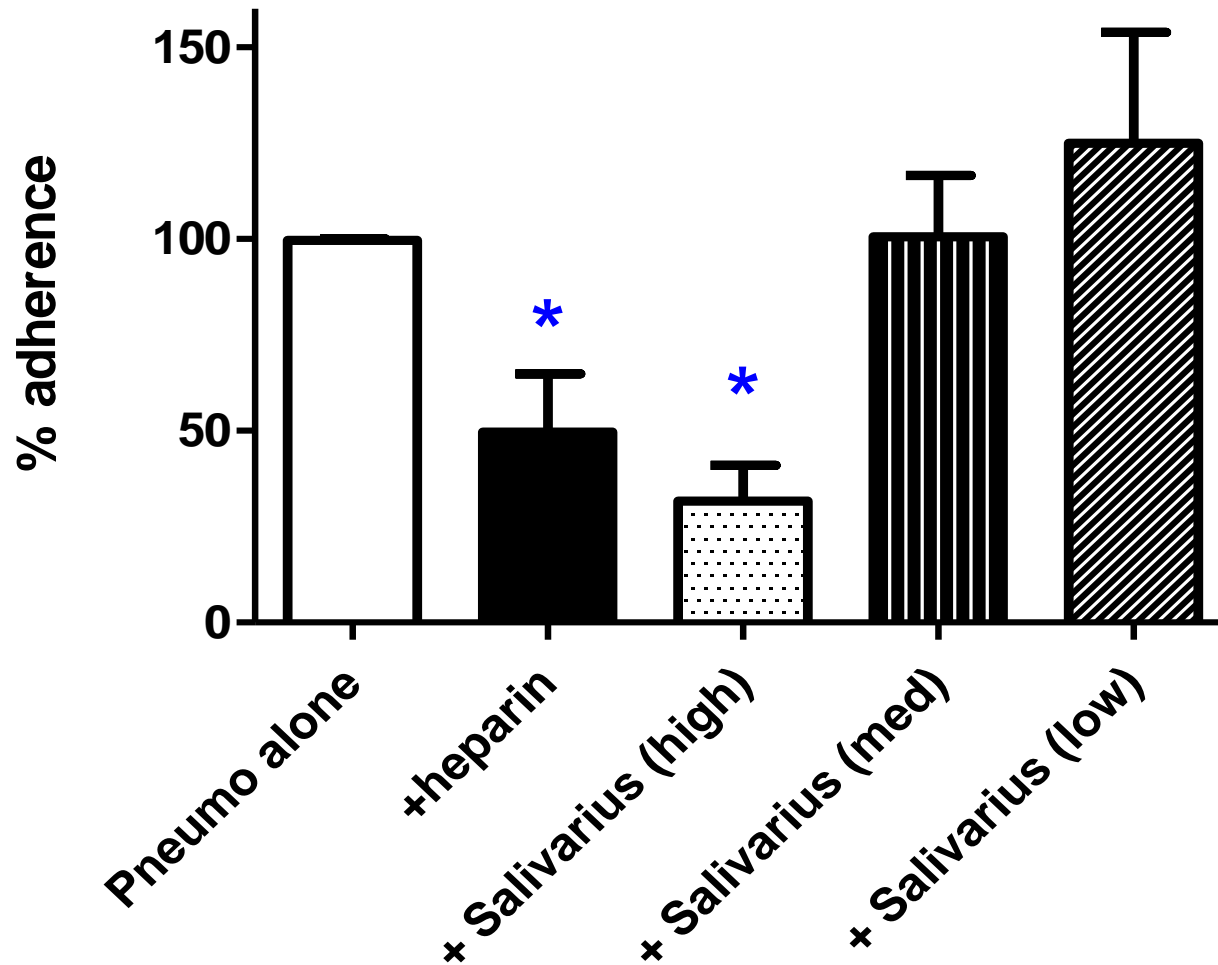
Results of *in vitro* assays

Pre-administration



n=4; * indicates $p < 0.05$ when compared to pneumo alone which was normalised to 100% for each assay (Students t test)

Co-administration



n=3; * indicates $p < 0.05$ when compared to pneumo alone which was normalised to 100% for each assay (Students t test)

Conclusion

- Pre-administration and co-administration of *S. salivarius* show a dose dependent inhibition of *S. pneumonie* colonisation
- Pre-administration was more effective than co- administration
- Post administration – no effect
- On going experiments with 2 other serotypes

Future directions

- ***in vivo* mouse model experiment**
 - Does *S. salivarius* colonise the mouse?
 - Does *S. salivarius* inhibit the colonisation of *S. pneumoniae* ?
 - Does *S. salivarius* prevent OM in a mouse model?
- **Design a clinical trail**

Thank you...

- My supervisors – Prof Stephen O’Leary,
Prof Roy Robins Browne,
Dr Catherine Satzke,
Dr Odilia Wijburg,
Dr Paul Licciardi
- Dr Eileen Dunne, Zheng Quan Toh
- Prof John Tagg and his colleagues
- Graeme Clark Foundation
- Hugh Noel Puckel Scholarship
- The University of Melbourne

Clinical trial

- Does *S. salivarius* inhibit the colonisation of *S. pneumoniae* in the nasopharynx ?
- Does *S. salivarius* prevent OM?
- If *S. salivarius* is effective in OM, then
 - mode of administration
 - dose of administration
 - time of administration

Statistical Analysis

- Results were compared to pneumococcal adherence in wells containing *S. pneumoniae* alone, normalised to 100%. The student's t-test was used to evaluate differences % adherence between test conditions and *S. pneumoniae* alone. P values < 0.05 were considered statistically significant and are indicated with an asterisk.

qPCR

- Lyta gene specific for pneumococcus used

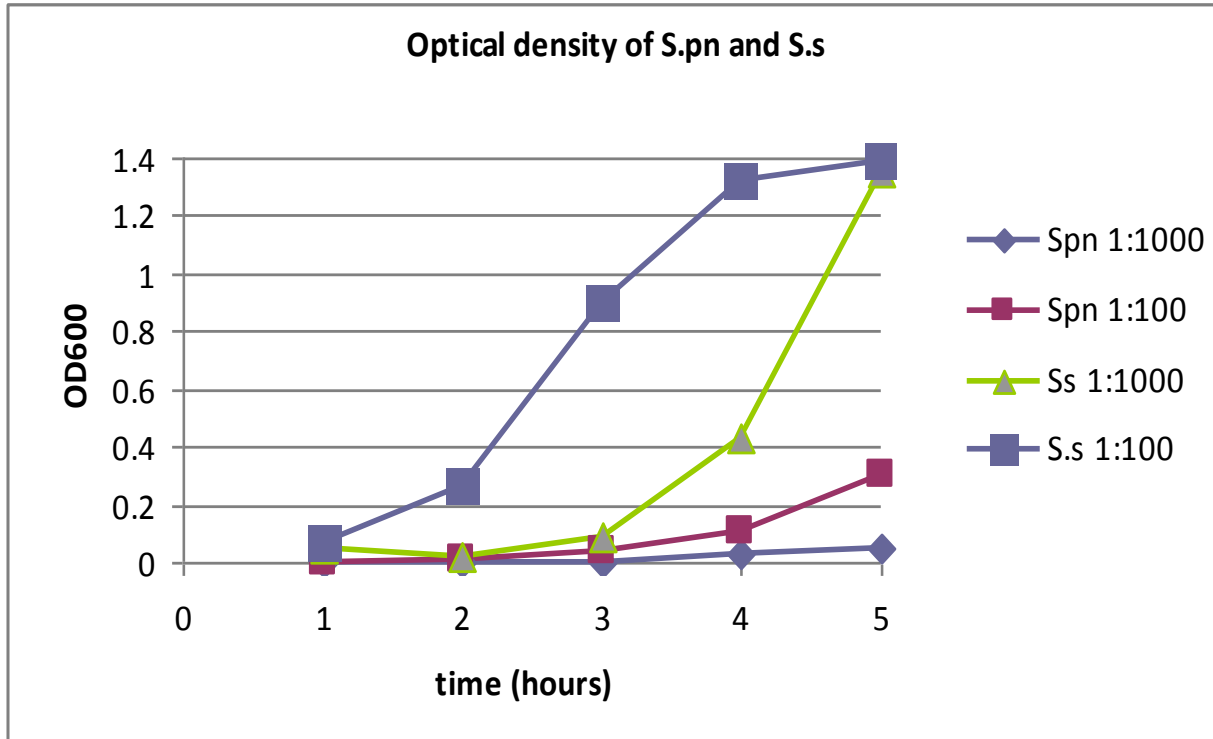


Figure 1. Growth of *S. pneumoniae* and *S. salivarius* over time as measured by optical density (OD₆₀₀).

Comparing qPCR to serial dilution and plating

	qPCR		Serial dilution & plating	
	CFU/ml PMP843	% colonisation	CFU/ml PMP 843	% colonisation
Pnc	1.1×10^7	100	9.1×10^6	100
Pnc + Heparin	1.5×10^6	14	7.6×10^5	8
Pnc + Ss (high)	6.1×10^5	6	3.2×10^5	4
Pnc + Ss (medium)	3.6×10^6	33	2.7×10^6	29
Pnc + Ss (low)	9.8×10^6	92	8.2×10^6	90

In vitro colonisation of *S. pneumoniae* (pre)

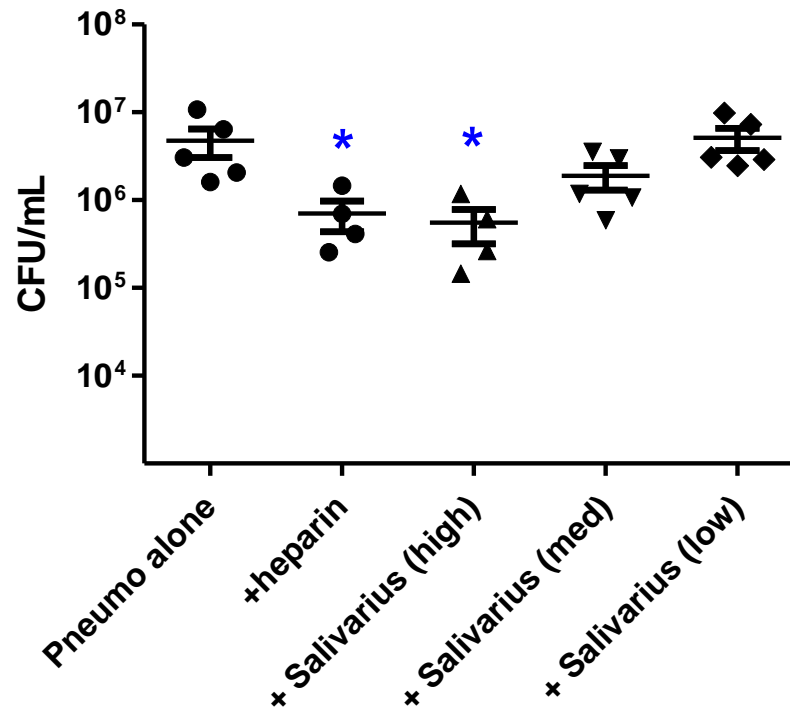


Figure 3. Colonisation of *S. pneumoniae* (CFU/mL) on Hep-2 cells when incubated alone, with heparin, or following pre-administration of high (5-10 *S. salivarius*: 1 *S. pneumoniae*), medium (1 *S. salivarius*: 1 *S. pneumoniae*), or low (1 *S. salivarius*: 5-10 *S. pneumoniae*) *S. salivarius*. * indicates $p < 0.05$ when compared to *S. pneumoniae* alone (Mann Whitney test).

In vitro colonisation of *S. pneumoniae* (co)

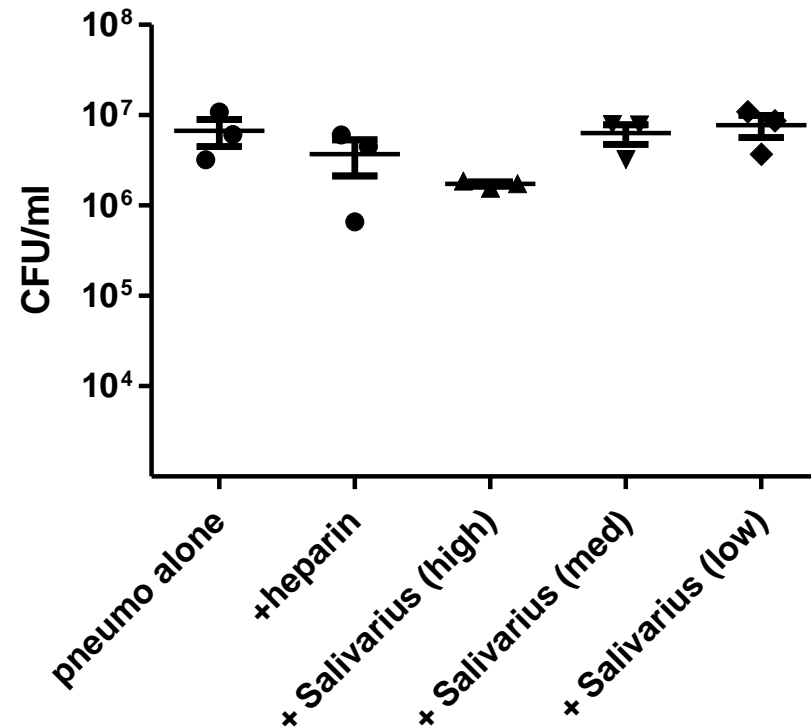


Figure 5. Colonisation of *S. pneumoniae* (CFU/mL) on Hep-2 cells when incubated alone, with heparin, or following co-administration of high (5-10 *S. salivarius*: 1 *S. pneumoniae*), medium (1 *S. salivarius*: 1 *S. pneumoniae*), or low (1 *S. salivarius*: 5-10 *S. pneumoniae*) *S. salivarius*.